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Medical Policy

Circulating Tumor DNA and Circulating Tumor Cells for Cancer Management (Liquid Biopsy)

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Policy Number: 797

BCBSA Reference Number: 2.04.141

Related Policies

Biomarkers for the Diagnosis and Cancer Risk Assessment of Prostate Cancer, #336

Policy¹

Commercial Members: Managed Care (HMO and POS), PPO, and Indemnity

Plasma-based comprehensive somatic genomic profiling testing (CGP) using Guardant360® for patients with Stage IIIB/IV non-small cell lung cancer (NSCLC) is considered <u>MEDICALLY NECESSARY</u> when the following criteria have been met:

Diagnosis:

- When tissue-based CGP is infeasible (i.e., quantity not sufficient for tissue-based CGP or invasive biopsy is medically contraindicated), AND
- When prior results for ALL of the following tests are not available:
 - o EGFR single nucleotide variants (SNVs) and insertions and deletions (indels)
 - ALK and ROS1 rearrangements
 - PDL1 expression.

Progression:

- Patients progressing on or after chemotherapy or immunotherapy who have never been tested for EGFR SNVs and indels, and ALK and ROS1 rearrangements, and for whom tissue-based CGP is infeasible (i.e., quantity not sufficient for tissue-based CGP), OR
- For patients progressing on EGFR tyrosine kinase inhibitors (TKIs).

If no genetic alteration is detected by Guardant360®, or if circulating tumor DNA (ctDNA) is insufficient/not detected, tissue-based genotyping should be considered.

Other plasma-based CGP tests are considered INVESTIGATIONAL.

CGP and the use of circulating tumor DNA is considered **INVESTIGATIONAL** for all other indications.

The use of circulating tumor cells is considered **INVESTIGATIONAL** for all indications.

Prior Authorization Information

Inpatient

 For services described in this policy, precertification/preauthorization <u>IS REQUIRED</u> for all products if the procedure is performed <u>inpatient</u>.

Outpatient

• For services described in this policy, see below for products where prior authorization <u>might be</u> required if the procedure is performed outpatient.

	Outpatient
Commercial Managed Care (HMO and POS)	The requirements of BCBSMA Genetic Testing Management Program require prior authorization via AIM Specialty Health. These requirements are member-specific:
Commercial PPO and	member specific.
Indemnity	Please verify member eligibility and requirements through Online Services by logging onto <u>Provider Central</u> . Refer to our <u>Quick Tip</u> for an overview of pre-certification and prior authorization requirements. Ordering clinicians should request prior authorization from <u>AIM Specialty Health</u> . <u>AIM's ProviderPortal</u> SM registration is required or call 1-866-745-1783 (when applicable).

CPT Codes / HCPCS Codes / ICD Codes

Inclusion or exclusion of a code does not constitute or imply member coverage or provider reimbursement. Please refer to the member's contract benefits in effect at the time of service to determine coverage or non-coverage as it applies to an individual member.

Providers should report all services using the most up-to-date industry-standard procedure, revenue, and diagnosis codes, including modifiers where applicable.

The following codes are included below for informational purposes only; this is not an all-inclusive list.

The above <u>medical necessity criteria MUST</u> be met for the following codes to be covered for Commercial Members: Managed Care (HMO and POS), PPO, and Indemnity:

CPT Codes

CPT codes:	Code Description
	Targeted genomic sequence analysis panel, solid organ neoplasm, DNA analysis, and
	RNA analysis when performed, 5-50 genes (eg, ALK, BRAF, CDKN2A, EGFR, ERBB2, KIT, KRAS, NRAS, MET, PDGFRA, PDGFRB, PGR, PIK3CA, PTEN, RET), interrogation
81445	for sequence variants and copy number variants or rearrangements, if performed
	Targeted genomic sequence analysis panel, hematolymphoid neoplasm or disorder, DNA
	analysis, and RNA analysis when performed, 5-50 genes (eg, BRAF, CEBPA, DNMT3A,
	EZH2, FLT3, IDH1, IDH2, JAK2, KRAS, KIT, MLL, NRAS, NPM1, NOTCH1), interrogation
	for sequence variants, and copy number variants or rearrangements, or isoform
81450	expression or mRNA expression levels, if performed
	Targeted genomic sequence analysis panel, solid organ or hematolymphoid neoplasm,
	DNA analysis, and RNA analysis when performed, 51 or greater genes (eg, ALK, BRAF,
	CDKN2A, CEBPA, DNMT3A, EGFR, ERBB2, EZH2, FLT3, IDH1, IDH2, JAK2, KIT,
81455	KRAS, MLL, NPM1, NRAS, MET, NOTCH1, PDGFRA, PDGFRB, PGR, PIK3CA, PTEN,

RET), interrogation for sequence variants and copy number variants or rearrangements, if
performed

Description

Liquid biopsy refers to analysis of circulating tumor DNA (ctDNA) or circulating tumor cells (CTCs) as a method of noninvasively characterizing tumors and tumor genome from the peripheral blood.

Circulating Tumor DNA

Normal and tumor cells release small fragments of DNA into the blood, which is referred to as cell-free DNA (cfDNA). cfDNA from nonmalignant cells is released by apoptosis. Most cell-free tumor DNA is derived from apoptotic and/or necrotic tumor cells, either from the primary tumor, metastases, or CTCs. Unlike apoptosis, necrosis is considered a pathologic process, and generates larger DNA fragments due to an incomplete and random digestion of genomic DNA. The length or integrity of the circulating DNA can potentially distinguish between apoptotic and necrotic origin. ctDNA can be used for genomic characterization of the tumor.

Circulating Tumor Cells

Intact CTCs are released from a primary tumor and/or a metastatic site into the bloodstream. The half-life of a CTC in the bloodstream is short (1-2 hours), and CTCs are cleared through extravasation into secondary organs. Most assays detect CTCs through the use of surface epithelial markers such as EpCAM and cytokeratins. The primary reason for in detecting CTCs is prognostic, through quantification of circulating levels.

Technologies for Detecting ctDNA and CTCs

Detection of ctDNA is challenging because ctDNA is diluted by nonmalignant circulating DNA and usually represents a small fraction (<1%) of total cfDNA. Therefore, more sensitive methods than the standard sequencing approaches (eg, Sanger sequencing) are needed.

Highly sensitive and specific methods have been developed to detect ctDNA, for both single-nucleotide mutations (eg BEAMing [which combines emulsion polymerase chain reaction [PCR] with magnetic beads and flow cytometry] and digital PCR) and copy-number changes. Digital genomic technologies allow for enumeration of rare mutant variants in complex mixtures of DNA.

Approaches to detecting ctDNA can be considered targeted, which h includes the analysis of known genetic mutations from the primary tumor in a small set of frequently occurring driver mutations, which can impact therapy decisions (eg, *EGFR* and *ALK* in non-small-cell lung cancer), or untargeted without knowledge of specific mutations present in the primary tumor, and include array comparative genomic hybridization, next-generation sequencing, and whole exome and genome sequencing.

CTC assays usually start with an enrichment step that increases the concentration of CTCs, either on the basis of biologic properties (expression of protein markers) or physical properties (size, density, electric charge). CTCs can then be detected using immunologic, molecular, or functional assays.

Summary

Circulating tumor DNA (ctDNA) and circulating tumor cells (CTCs) in peripheral blood, referred to as "liquid biopsy," potentially offer a noninvasive alternative to tissue biopsy for therapeutic decisions and clinical prognosis in patients with cancer.

For individuals who have cancer who receive molecular characterization of tumor using ctDNA, the evidence includes case series and systematic reviews of these case series. Relevant outcomes are overall survival, disease-specific survival, test accuracy and validity, morbid events, and medication use. Ultrasensitive methods to detect mutations from ctDNA have been developed, but there is limited evidence on the analytic validity of these methods. There is a need for further optimization and standardization of testing methods. Clinical validity consists of case series that report correlations between mutations detected in ctDNA with mutations detected in tumor tissue. Results have shown variable results for clinical sensitivity. Although some reports have suggested that clinical sensitivity may

be high, this finding has not been consistent. Published studies have consistently reported high clinical specificity; however, most study populations are small and heterogeneous, and it is not known to what degree mutations detected by ctDNA are representative of the primary tumor. Published studies reporting clinical outcomes and/or clinical utility are lacking. However, specifically for ctDNA in non-small cell lung cancer (NSCLC), the evidence supports improved health outcomes at tumor progression and at diagnosis if tissue sample is unobtainable.

For individuals who have cancer or are at high risk of developing cancer who receive identification and quantification of CTCs, the evidence includes case series and meta-analyses of these case series. Relevant outcomes are overall survival, disease-specific survival, and test accuracy and test validity.

Published data on analytic validity are lacking. Most of the literature consists of reports of levels of CTCs and cancer prognosis, and have shown a correlation with survival in certain cancer types. However, the cutoff levels that should be used to signal a change in patient management are unknown, and there are no studies showing clinical utility and improved patient outcomes. The evidence is insufficient to determine the effects of the technology on health outcomes. If a separate evidence review exists, then conclusions reached there supersede conclusions in this review.

Policy History

Date	Action
1/2021	Medicare information removed. See MP #132 Medicare Advantage Management for local coverage determination and national coverage determination reference. Clarified coding information.
6/2020	Policy reactivated. Prior authorization clarified. Prior authorization is required through AIM Specialty Health. Effective 6/8/2020.
10/2017	Clarified coding information.
9/1/2017	New medically necessary indications described. Clarified coding information. Effective 9/1/2017.
10/2016	New medical policy describing investigational indications. Effective 10/1/2016.

Information Pertaining to All Blue Cross Blue Shield Medical Policies

Click on any of the following terms to access the relevant information:

Medical Policy Terms of Use

Managed Care Guidelines

Indemnity/PPO Guidelines

Clinical Exception Process

Medical Technology Assessment Guidelines

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Endnotes

¹ Based on MPRM 2.04.141 and expert opinion.